

## Application of rice (*Oryza sativa* L.) suspension culture in studying senescence *in vitro* (II). Changes in DNA integrity

Ramin Hosseini<sup>\* #</sup>

Plant Research Group  
YAPO School of Biological Sciences  
University of Nottingham  
University Park Nottingham NG7 2RD  
United Kingdom  
E-mail: Raminh\_2001@Yahoo.com

Bernard J. Mulligan<sup>§</sup>

Plant Research Group  
School of Biological Sciences  
University of Nottingham  
University Park Nottingham NG7 2RD  
United Kingdom  
E-mail: Bernard.Mulligan@DG12.ccc.be

**Financial support:** Ministry of Science, Research and Technology of Iran.

**Keywords:** Programmed Cell Death (PCD), rice, senescence.

**Present address:** #Research Centre for Applied Sciences, Jahade Daneshgahi, Shahid Beheshti University, Evin, Tehran, Iran. Tel: 0098-21-2402097 (ext. 213). Fax: 0098-21-9403884. §European Commission, DG12, Office SDME 9/28, 200 Ruela Lai, 1049, Brussels, Belgium.

**Changes in the DNA content and organisation of senescing rice cell cultures (Taipei 309) were studied, using PCR and Southern blot analyses. A mitochondrial gene (*coxII*), a plastid gene (*psaA*) and a nuclear DNA maker (*RG64*) were analysed. The amplification of mitochondrial (mt), plastid and nuclear DNA produced the expected fragments, indicating that there were still some intact organelles and nuclei in the senescing rice cells. However, in plastid and nuclear DNA, changes in the number and size of the PCR products were observed. Southern blot analysis revealed that mt and nuclear DNA contents declined, but plastid DNA content remained relatively the same throughout the senescence.**

Development and differentiation in plants require that specific cells be eliminated by cell death mechanisms (Buckner et al. 1998). Programmed Cell Death (PCD) is the genetically controlled process of cell death, which occurs during development and in response to environmental cues (Greenberg, 1996). PCD has been well characterized in animals and is a genetically controlled process that facilitates the disassembly of the dying cell (Buckner et al. 1998). Animal cells undergoing PCD can display cytoplasmic condensation and shrinkage, blebbing of plasma membranes and nuclear membranes, chromatin condensation and marginalization, internucleosomal DNA

fragmentation and phagocytosis of the dying cell. When these morphological changes are observed, the cell death process is often termed apoptosis (Wyllie et al. 1980; Kressel and Groscurth, 1994). In plants, DNA degradation has been observed during certain developmental events, such as tracheary element development in pea (Mittler and Lam, 1995) in leaf senescence in rice (Sodmergen et al. 1989; Sodmergen et al. 1991), in cell senescence in *Arabidopsis* (Callard et al. 1996) and during the hypersensitive response to pathogen attack (Mittler and Lam, 1995; Wang et al. 1996; Mittler and Lam, 1997). In plants, changes in chloroplast ultrastructure (Greening et al. 1982; Meier and Lichtenthaler, 1982), autophagic activity (Aubert et al. 1996), accumulation of electron-dense granules in the cytoplasm, reduction in the number of mitochondrial cisternae and disorganisation of cell wall fibrillar material (Pareek et al. 1997) have been observed during senescence, environmental stress and induced starvation. Whether plant cells undergo apoptotic PCD, as defined in animal cell systems, remains an open question. In another paper (Hosseini and Mulligan, 2002) we showed growth kinetics, viability and protein content of Taipei 309 cells in a 4-week culture period. We also revealed that how acid and zinc dependent nucleases were activated in these cells. Now, a further step has been taken to discover whether the detected nuclease activity, has any effect on genomic and/or organellar DNAs, a phenomenon known to be a criterion of apoptosis in animal systems. The results

\* Corresponding author

are discussed in the context of current research on PCD in plants.

## Materials and Methods

Cell suspension cultures of the rice *cv.* Taipei 309 were grown in 100 ml conical flasks (Finch, 1991). One ml of settled cells was inoculated in a sterilised conical flask containing 22 ml of fresh AA2 medium (Ohira et al. 1973) and 7 ml of spent medium. The cells were grown on a rotary shaker at 120 rpm at 27°C ± 1 for 3 weeks post-subculture. Every week the cells were collected (three replicates) by sieving through 30 µm sieves and air-dried for 2 h on paper towels and stored at -20°C for future use.

### DNA extraction

Total DNA from Taipei 309 seedlings and 1-, 2- and 3-week old cells was isolated (Doyle and Doyle, 1987) and its concentration was determined (Sambrook et al. 1989).

### Primer design for PCR reactions

Three pairs of primers (20) bases were designed to allow amplification of nuclear, plastid and mitochondrial gene sequences (one of each). Published rice sequences were inspected to determine suitable sequences as follows:

- Mitochondrial gene *coxII* (Kao et al. 1984).
- Plastid gene *psaA* Gopher://megasun.bch.umonreal.ca:70/00/organelles/plstidswgeno.03
- Nuclear DNA marker *RG64* (on chromosome 3; Hittalmani et al. 1995).

The three primers (Table 1) were synthesised through the Department of Biochemistry, Queens Medical Centre, University of Nottingham and used for PCR reactions. The length of the gene/marker and size of expected amplified fragments are given in Table 2. Also PCR products were radiolabelled, using α [<sup>32</sup>P](dCTP; Amersham Life Science Ltd., Little Chalfont, UK ) and used as hybridisation probes.

### Conditions employed for PCR

Total DNA from Taipei 309 seedlings was used for the

optimisation of the PCR conditions. To optimise the PCR conditions, different concentrations (0.5 mM, 1 mM, 1.5 mM and 2 mM) of MgCl<sub>2</sub> in the reaction buffer and two annealing temperatures (45 and 50°C) were tested (see Results). After optimisation of PCR conditions, DNA samples from Taipei 309 seedlings and 1-, 2- and 3-week old Taipei 309 cell cultures were used for investigation of the integrity of *coxII*, *psaA* and *RG64*. The amplification of the desired DNA fragments was carried out based on the method described in Sambrook et al. 1989.

### Southern blot analysis

The amplified fragments of mt, plastid and nuclear DNA (see above) obtained by the PCR reactions from Taipei 309 seedlings were used as probes and Southern blot analysis of senescing cells of Taipei 309 was carried out; following the method described by Sambrook et al. 1989.

### Analysis and purification of the amplified DNA fragments

To analyse the PCR products, 6 µl of the reaction mixture was mixed with 1 µl of 6 x loading buffer [0.25% (w/v) bromophenol blue, 0.25% (w/v) xylene cyanol FF, 30% (v/v) glycerol] and electrophoresed on a 0.8% (w/v) agarose gel (5 x 6 cm; Sigma, type V) at 100 v for 30 min. The gel was stained with ethidium bromide (0.5 mg/l) for 30 min and the PCR products were visualised and photographed using a UV transilluminator (TM-20 model; UVP Ltd., California, USA). The amplified fragments were purified using a Prep-A-Gene kit according to the manufacturer's recommendation (Bio-Rad, Hemel Hempstead, UK).

## Results

### Changes in DNA integrity in senescing Taipei 309 cells, revealed by PCR

The integrity of mitochondrial, chloroplast and nuclear DNA during senescence of Taipei 309 cells was investigated using PCR. The DNA from Taipei 309 seedlings was used as a control, alongside DNA samples from 1-,2- and 3-week old Taipei 309 cells.

**Table1. Sequence of the three primers for rice mt and plastid genes and the nuclear DNA marker, RG64.**

Primer	Forward	Reverse
<i>CoxII</i>	CCTCACAATCGCTCTTTCTG	AACCTGTAGTTACCATAGCC
<i>PsaA</i>	ACTCCTACAGCACGTCCTTG	CAGTAGGCGGCAAAGTTGCT
<i>RG64</i>	CGGTGAGAGATATGCGTGC	TGTAGTACTACCTCCGTTCC

**Table 2.** The length of the gene/marker and the expected amplified fragment sizes.

Primer	Fragment size (bp)	Length of the gene/marker
<i>CoxII</i>	336	2048
<i>PsaA</i>	639	2312
<i>RG64</i>	524	1557

### Mitochondrial DNA (*coxII*)

PCR amplification of the 5' end of the rice *coxII* gene was carried out under the conditions described previously. Different concentrations of MgCl<sub>2</sub> were tested in the PCR. A 2mM concentration of MgCl<sub>2</sub> slightly inhibited the amplification of the *coxII* product, compared to the lower concentrations tested (Figure 1a). Therefore, 0.5 mM MgCl<sub>2</sub> was chosen as the optimum concentration. The expected fragment (336 bp) was amplified in all conditions used. In order to investigate the intactness of the mitochondrial DNA (*coxII*), PCR reactions were carried out using 0.05 µg of template DNA from Taipei 309 seedlings and from 1-, 2- and 3-week old cells. No differences were observed in number and size of the amplified products between samples (Figure 1b).

### Plastid DNA (*psaA*)

Different concentrations of MgCl<sub>2</sub> were tested to optimise the PCR conditions and 2 mM MgCl<sub>2</sub> was found to give the best amplification. The expected fragment (639 bp) was amplified but two smaller and diffuse (200 and 300 bp) products were also observed (Figure 2a). An attempt to restrict amplification of the extra bands by increasing the annealing temperature to 50°C failed (Figure 2b). In order to investigate the integrity of plastid DNA in the senescing Taipei 309 cells, a PCR reaction with DNA samples from Taipei 309 seedlings, 1-, 2- and 3-week old cells was run under optimal conditions. The 639 bp was amplified in all samples (Figure 2c) but the two diffuse bands were less obvious in the PCR product from 2-week-old cells and barely detectable in the PCR product from 3-week-old cells (Figure 2c).

### Nuclear DNA (*RG64*)

The optimum MgCl<sub>2</sub> concentration was found to be 1 mM (Figure 3a). In addition to the expected product (524 bp), two smaller fragments (about 220 and 340 bp) were also observed (Figure 3a). The same products were amplified when the primer annealing temperature was increased to 50°C (Figure 3b). Changes in nuclear DNA (*RG64*) integrity were investigated by running a PCR reaction using DNA templates from Taipei 309 seedlings, 1-, 2- and 3-week old cells under optimal conditions. In addition to the expected product (524 bp), as observed when using DNA from cultured cells, three other products (220 bp, 340 bp,

1000 bp) were amplified in the Taipei 309 seedling sample (Figure 3c). The abundance of the 1000 bp product was greater when PCRs were carried out using DNA from 2- and 3-week-old cells compared to seedlings and 1-week-old cells. The other amplified DNA products (220 and 340 bp) from 1- and 2- week- old cells were similar in abundance to those produced from seedlings, though the 524 bp and 340 bp products were reduced in 3-week-old cells (Figure 3c). In the amplified products from 3-week-old cells, a faint extra band of about 1600 bp was also produced (Figure 3c).

### Southern blot analysis of mitochondrial, plastid and nuclear DNA of senescing cells of Taipei 309

Southern blot analysis of DNA from Taipei 309 cells was carried out in order to detect any changes in the organellar and/or nuclear DNA as the cells senesced. PCR amplified fragments of a mitochondrial gene, *coxII*, a plastid gene, *psaA*, and a nuclear DNA marker, *RG64* were radiolabelled and used as probes (see above).

### Mitochondrial DNA (*coxII*)

Total DNA samples (5 µg) from Taipei 309 seedlings, 1-, 2- and 3-week old cells were digested with *Bam*HI (Figure 4a) and hybridised to a PCR fragment corresponding to the 5' end of the rice mitochondrial gene, *coxII* (see above). In seedlings and 1-week old cells, the probe hybridised to one fragment (7.2 kb). In 2-week-old cells, however, 2 extra bands of 3.5 and 1.5 kb were also observed. The intensity of the 7.2 kb signal decreased with cell age and in 3-week-old cells, only a very weak hybridisation to the 7.2 kb fragment was observed (Figure 4b). It was clear that DNA content declined progressively in 1- to 3-week-old cells (Figure 4b).

### Plastid DNA (*psaA*)

Total DNA (5 µg) from Taipei 309 seedlings, 1-, 2- and 3-week-old cells was digested with *Bam*HI (Figure 5a) and hybridised to a PCR fragment of the rice chloroplast gene *psaA*. In the seedlings and cell samples there was a strong 2.2 kb signal whose intensity remained the same regardless of cell age. In addition to this band, in 2-week-old cells, four very weak signals of 1.9, 1.7, 1.6 and 1.3 kb were observed. In 3-week-old cells, the hybridisation pattern was similar to that of 2-week-old cells but the 1.9, 1.7, 1.6 and 1.3 kb signals were unclear (Figure 5b).

## Nuclear DNA (RG64)

For nuclear DNA analysis, total DNA (5 µg) was digested with *Bam*HI (Figure 6a), and hybridised to a PCR fragment of *RG64*, a rice nuclear DNA marker. All samples from seedlings and cells showed 6 signals, with sizes of 9.4, 7.2, 2.9, 2.4, 1.5 and 1.3 kb. The strongest signals were obtained with seedling DNA but the signals became progressively weaker in 1-, 2- and 3-week-old cells (Figure 6b).

## Discussion

### Changes in DNA integrity of senescing rice cells

Senescence is a controlled sequence of events comprising the final stage of development (Greening et al. 1982; Guamet and Giannibelli, 1994; Smart, 1994). The existence of senescence-related genes provides the most direct evidence for senescence being controlled by gene expression (Thomas and Howarth, 2000) as this has been recently confirmed by Hinderhofer and Zentgraf, 2001.

DNA degradation is one of the events occurring during senescence (Sodmergen et al. 1989; Sodmergen et al. 1991). DNA degradation has been reported to occur during other plant developmental stages (Blank and McKeon, 1989; Mittler and Lam, 1995; Callard et al. 1996; Greenberg, 1996; Ryerson and Heath, 1996; Wang et al. 1996).

In the present study, in order to find out any changes in DNA related to cell senescence, the integrity of nuclear, mt and plastid DNA of Taipei 309 cells was investigated by PCR and Southern blot analysis.

### mtDNA

As expected, use of the *coxII* primers in PCRs, with total DNA from rice seedlings as substrate, produced a 336 bp amplified DNA fragment. The same sized fragment was produced when total DNA from 1-3 week old suspension cultures was employed as substrate. This result suggested that some intact mitochondrial DNA was still present even in 3-week-old cultures. Under the PCR conditions employed, it was not possible to conclude whether mitochondrial DNA (per cell or per g.f.wt. cells) decreased from week 1 to week 3, neither was it possible on the basis of the PCR data alone to assess the integrity of the mtDNA. However, the Southern hybridisation data suggested that the proportion of total DNA represented by the mt genome decreased with cell age and at the same time changes in mt DNA occurred.

Aubert et al. 1996 suggested that in plants, signal(s) giving rise to the autophagic process originate(s) from the decline of supply of respiratory substrates to the mitochondria. It

seems that in plant senescence, the role of mitochondrial DNA alterations has not been fully investigated, even though the mitochondria have been found intact until the late stages of senescence (Smart, 1994). The data presented here suggested that mtDNA alterations were occurring in the rice cells as they senesced.

### Plastid DNA

*psaA* and *psaB* genes encode for the two polypeptides of P700 chlorophyll *a* protein complex of photosystem I reaction centre. These two genes are located adjacent to each other in the large single copy region of the circular plastid genome in higher plants (Chen et al. 1992). Primers for plastid DNA (*psaA*) showed several amplified bands in PCR, whereas since *psaA* is a single copy sequence (Hiratsuka et al. 1989), only one band was expected. Increasing the annealing temperature to 50°C failed to solve the problem of the extra bands. It may suggest that there are homologous sequences to the investigated sequence (*psaA*) in the rice genome. The expected *psaA* band was amplified in seedlings and cell samples. In seedlings, 1- and 2-week old cells the two diffuse bands (200 and 300 bp) were present but in the PCR product from 3-week-old cultures, the intensity of the two extra bands reduced. Thus, as judged by PCR, plastid DNA showed some differences in 2- and 3-week-old cells compared to the seedlings and 1-week-old cells. These findings may suggest that subtle changes had taken place in plastid DNA of the senescing rice cells. Interestingly, however, Southern blots showed that the abundance of plastid DNA was only slightly affected by cell age. There are contradictory reports about chloroplast integrity during senescence. Wittenbach et al. 1982 reported that chloroplasts could be isolated from protoplasts prepared from dark-induced senescent wheat leaves from which up to 80% of chlorophyll and protein had been lost. Consistent with the results presented here, however, Jones and Dangel, 1996 mentioned that plastids appear to be the last organelles to be degraded. Greening et al. 1982 reported the loss of plastid integrity, while Sodmergen et al. 1989 and Sodmergen et al. 1991 reported a decline in cpDNA at early stages of senescence. However, in this research, it was observed that in rice cells, the quantity of plastid DNA remained the same throughout the process of senescence; even though qualitative alterations of plastid DNA were observed.

Bate et al. 1990, reported that during senescence in bean plants, D-1 the product of *psaA*, a PSII (photosystem II) protein, continued to be actively synthesised despite a 9-fold decrease in RNA synthesis and the reduced synthesis of all major thylakoid proteins. They also found that in senescent bean, the synthesis of *psaA* RNA was unaltered during senescence. In the present study, Southern blot analysis of *psaA* in senescing Taipei309 cells revealed that this gene remained intact up to very late stages of

senescence. Significant transcript levels of *psaA* and *psaB* have also been observed in dark grown barley seedlings (Kreus et al. 1986).

### Nuclear DNA

When the amplification of *RG64* from seedlings and cells was carried out, several extra PCR products were detected in addition to the expected product (524 bp). However, in 3-week-old cells the 524 bp band was reduced in abundance and became diffuse but one of the extra bands (1000 bp product) intensified and another extra band about 1.6 kb was produced. Nuclear DNA samples showed the same hybridisation pattern to the *RG64* probe but a decrease in DNA quantity with cell age was observed.

Guarente, 1996 suggests that changes in chromosomal structure or function may be a key determinant of ageing. In plants, nuclei remain intact up to the late stages of senescence (Smart, 1994). It has been suggested that nuclear genes control the degradation of chloroplasts (Guiamet and Giannibelli, 1994).

### Concluding Remarks

The PCR assay revealed no alteration to mtDNA in ageing rice cells, whereas Southern blot showed 2 extra signals in 2-week-old cells and a progressive decrease in mtDNA content. Plastid DNA had undergone some rearrangements, which was shown by both PCR and Southern blot analysis, with no decrease in its quantity. Thus, qualitative and quantitative changes were detected in nuclear DNA by PCR and Southern blot analysis, respectively. Nuclear and mtDNA content of the senescing rice cells declined. However, plastid DNA content showed subtle changes. The decrease in DNA content may imply an increase in nuclease activity in senescing cells. DNA degradation, a feature of animal cell death, apoptosis, was observed in senescing rice cells. However, the changes in DNA content and integrity should be more fully investigated using a number of genes. Following the investigation of changes at DNA level, discovering changes at transcription and translation levels would also be beneficial.

### References

AUBERT, S.; GOUT, E.; BLIGNY, R.; MARTY-MAZARS, D.; BARRIEU, F.; ALABOUVETTE, J.; MARTY, F. and DOUCE, R. Ultrastructural and biochemical characterization of autophagy in higher plant cells subjected to carbon deprivation: control by the supply of mitochondria with respiratory substrates. *Journal of Cell Biology*, 1996, no. 133, p. 1251-1263.

BATE, N.J.; STRAUS, N.A. and THOMPSON, J.E. Expression of chloroplast photosynthesis genes during leaf

senescence. *Physiologia Plantarum*, 1990, no. 80, p. 217-225.

BLANK, A. and MCKEON, T.A. Single-strand-preferring nuclease activity in wheat leaves is increased in senescence and is negatively photoregulated. *Proceedings of the National Academy of Sciences of the United States of America*, 1989, no. 86, p. 3169-3173.

BUCKNER, B.; JANICK-BUCKNER, D. and GRAY, J. Cell death mechanisms in maize. *Trends in Plant Sciences*, 1998, no. 3, p. 218-223.

CALLARD, D.; AXELOS, M. and MAZZOLINI, L. Novel molecular markers for late phases of the growth cycle of *Arabidopsis thaliana* cell-suspension cultures are expressed during organ senescence. *Plant Physiology*, 1996, no. 112, p. 705-715.

CHEN, S.C.; CHENG, M.; CHUNG, K.; YU, N. and CHEN, M. Expression of the rice chloroplast *psaA-psaB-rps 14* gene cluster. *Plant Science*, 1992, no. 81, p. 93-102.

DOYLE, J.J. and DOYLE, J.L. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical Bulletin*, 1987, no. 19, p. 11-15.

FINCH, R. Development of protoplast systems for genetic manipulation of rice. Ph.D. Thesis, University of Nottingham, Nottingham, UK. 1991.

GREENBERG, J.T. Programmed cell death: a way of life for plants. *Proceedings of the National Academy of Sciences of the United States of America*, 1996, no. 93, p. 12094-12097.

GREENING, M.T.; BUTTERFIELD, F.J. and HARRIS, N. Chloroplast ultrastructure during senescence and regreening of flax cotyledons. *New Phytologist*, 1982, no. 92, p. 279-285.

GUARENTE, L. Do changes in chromosomes cause ageing? *Cell*, 1996, no. 86, p. 9-12.

GUIAMET, J.J. and GIANNIBELLI, M.C. Inhibition of the degradation of chloroplast membranes during senescence in nuclear 'stay green' mutants of soybean. *Physiologia Plantarum*, 1994, no. 91, p. 395-402.

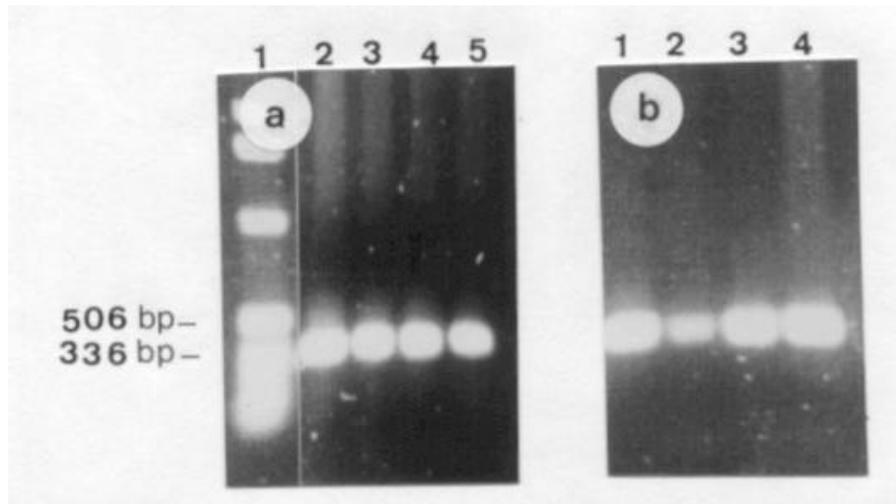
HINDERHOFER, K. and ZENTGRAF, U. Identification of a transcription factor specifically expressed at the onset of leaf senescence. *Planta*, 2001, no. 213, p. 469-477.

HIRATSUKA, J.; SHIMADA, H.; WHITTIER, R.; ISHIBASHI, T.; SAKAMOTO, M.; MORI, M.; KONDO, C.; HONJI, Y.; SUN, C.; MENG, B.; LI, Y.; KANNO, A.;

- NISHIZAMA, Y.; HIRAI, A.; SHINOZAKI, K. and SUGIURA, M. The complete sequence of the rice (*Oryza sativa*) chloroplast genome: intermolecular recombination between distinct tRNA genes account for a major plastid DNA inversion during evolution of the cereals. *Molecular and General Genetics*, 1989, no. 217, p. 185-194.
- HITTALMANI, S.; FOOLAD, M.R.; MEW, T.; RODRIGUEZ, R.L. and HUANG, N. Development of a PCR-based marker to identify rice blast resistance gene, *Pi-2(t)*, in a segregating population. *Theoretical and Applied Genetics*, 1995, no. 91, p. 9-14.
- HOSSEINI, Ramin and MULLIGAN, Bernard J. The application of rice (*Oryza sativa* L.) suspension cultures in studying senescence (I). Single Strand-Prefering Nuclease activity. *EJB Electronic Journal of Biotechnology* [on line]. 15 April 2002, vol. 5, no. 1 [cited 15 April 2002]. Available from Internet: <http://www.ejb.org/content/vol5/issue1/full/8/>. SSN 07117-3458.
- JONES, A.M. and DANGL, J.F. Logim at Styx: programmed cell death in plants. *Trends in Plant Sciences*, 1996, no.1, p. 114-119.
- KAO, T.; MOON, E. and WU, R. Cytochrome oxidase subunit II gene of rice has an insertion sequence within the intron. *Nucleic Acids Research*, 1984, no. 12, p. 7305-7315.
- KRESSEL, M. and GROSCURTH, P. Distinction of apoptotic and necrotic cell death by *in situ* labeling of fragmented DNA. *Cell Tissue Research*, 1994, no. 278, p. 549-556.
- KREUS, K.; DEHESH, K. and APEL, K. The light-dependent accumulation of the P700 chlorophyll *a* protein of the photosystem I reaction center in barley. Evidence for translational control. *European Journal of Biochemistry*, 1986, no. 159, p. 459-467.
- MEIER, D. and LICHTENTHALER, H.K. Special senescence stages in chloroplast ultrastructure of radish seedlings induced by the photosystem II-herbicide bentazon. *Protoplasma*, 1982, no. 110, p. 138-142.
- MITTLER, R. and LAM, E. Characterization of nuclease activities and DNA fragmentation induced upon hypersensitive response cell death and mechanical stress. *Plant Molecular Biology*, 1997, no. 34, p. 209-221.
- MITTLER, R. and LAM, E. *In situ* detection of nDNA fragmentation during the differentiation of tracheary elements in higher plants. *Plant Physiology*, 1995, no. 108, p. 489-493.
- OHIRA, K.; OJIMA, K. and FUJIWARA, A. Studies on the nutrition of rice cell culture I. A simple, defined medium for rapid growth in suspension culture. *Plant Cell Physiology*, 1973, no. 14, p. 1113-1121.
- PAREEK, A.; SINGLA, S.L. and GROOVER, A. Short-term salinity and high temperature stress-associated ultrastructural alterations in young leaf cells of *Oryza sativa* L. *Annals of Botany*, 1997, no. 80, p. 629-639.
- RYERSON, D.E. and HEATH, M.C. Cleavage of nuclear DNA into oligonucleosomal fragments during cell death induced by fungal infection or by abiotic treatments. *The Plant Cell*, 1996, no. 8, p. 393-402.
- SAMBROOK, J.; FRITSCH, E.F. and MANIATIS, T. *Molecular Cloning*. Cold Spring Harbor Laboratory Press, New York, USA, 1989. 1659 p.
- SMART, C.M. Gene expression during leaf senescence. *New Phytologist*, 1994, no. 126, p. 419-448.
- SODMERGEN KAWANO, S.; TANO, S. and KUROIWA, T. Degradation of chloroplast DNA in second leaves of rice (*Oryza sativa*) before leaf yellowing. *Protoplasma*, 1991, no. 160, p. 89-98.
- SODMERGEN KAWANO, S.; TANO, S. and KUROIWA, T. Preferential digestion of chloroplast nuclei (nucleoids) during senescence of the coleoptile of *Oryza sativa*. *Protoplasma*, 1989, no. 152, p. 65-68.
- THOMAS, H. and HOWARTH, C.J. Five ways to stay green. *Journal of Experimental Botany*, 2000, no. 51, p. 329-337.
- WANG, H.; JUAN, LI.; BOSTOCK, R.M. and GILCHRIST, D.C. Apoptosis: a functional paradigm for programmed plant cell death induced by a host-selective phytotoxin and invoked during development. *The Plant Cell*, 1996, no. 8, p. 375-391.
- WITTENBACH, V.A.; LIN, W. and HEBERT, R.R. Vacuolar localisation of proteases and degradation of chloroplasts in mesophyll protoplasts from senescing primary wheat leaves. *Plant Physiology*, 1982, no. 69, p. 98-102.
- WYLLIE, A.H.; KERR, J.F.R. and CURRIES, A.R. Cell death: the significance of apoptosis. *International Review of Cytology*, 1980, no. 68, p. 251-306.

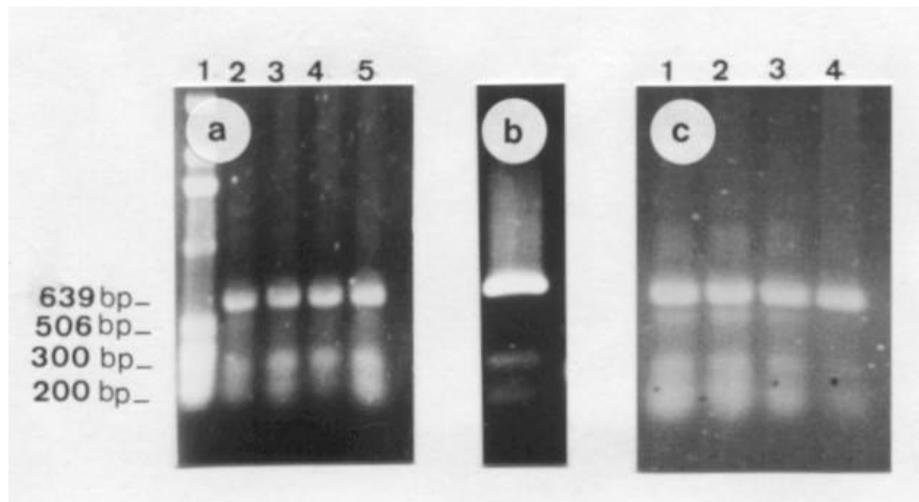
## APPENDIX

### Figures



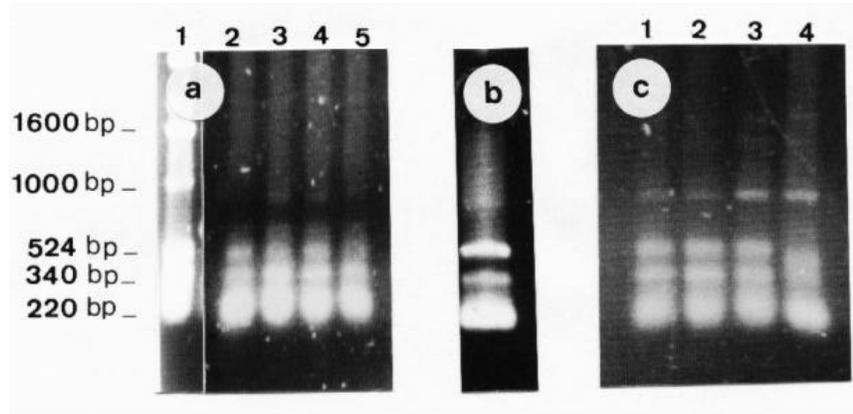
**Figure 1.**

- PCR amplification of the 5' end of the mitochondrial gene *coxII* using total DNA from Taipei 309 seedlings as substrate. Different concentrations of  $MgCl_2$  were used to optimise the PCR conditions. Lane 1, DNA marker, lane 2, 0.5 mM  $MgCl_2$ , lane 3, 1 mM  $MgCl_2$ , lane 4, 1.5 mM  $MgCl_2$  and lane 5, 2 mM  $MgCl_2$ .
- PCR amplification of the 5' end of *coxII* under optimal conditions. Lane 1, DNA sample from Taipei 309 seedlings, lanes 2-4 DNA samples from 1-, 2- and 3-week-old cells.



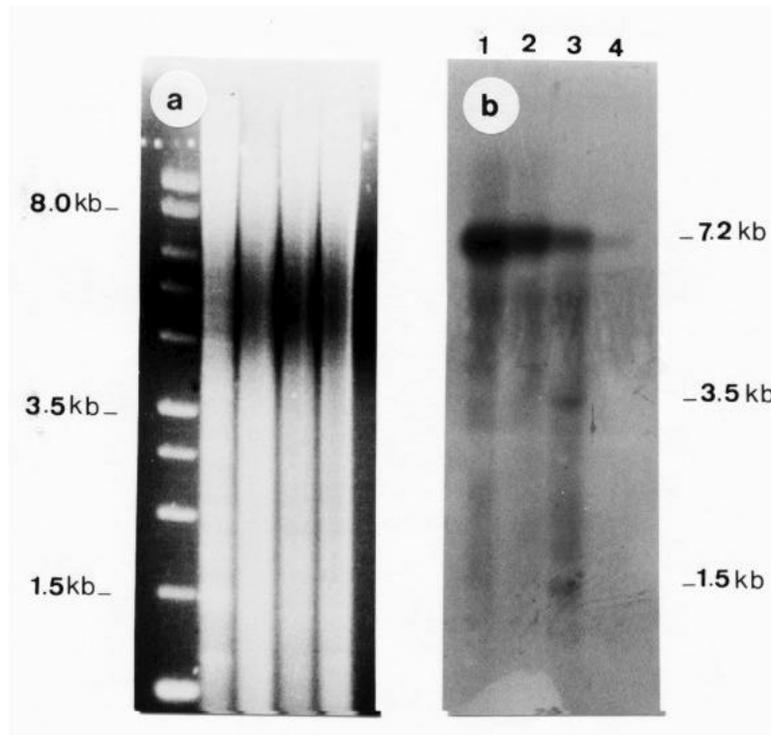
**Figure 2.**

- Amplification of the plastid gene *psaA* from Taipei 309 seedlings using different concentrations of  $MgCl_2$ . Lane 1, DNA marker, lane 2, 0.5 mM  $MgCl_2$ , lane 3, 1 mM  $MgCl_2$ , lane 4, 1.5 mM  $MgCl_2$  and lane 5, 2 mM, all at 45°C.
- The annealing temperature was raised to 50°C to remove the extra bands.
- Amplification of *psaA* from Taipei 309 seedlings and cells. Lane 1, DNA sample from Taipei 309 seedlings, lanes 2-4 DNA samples from 1-, 2- and 3-week-old cells.



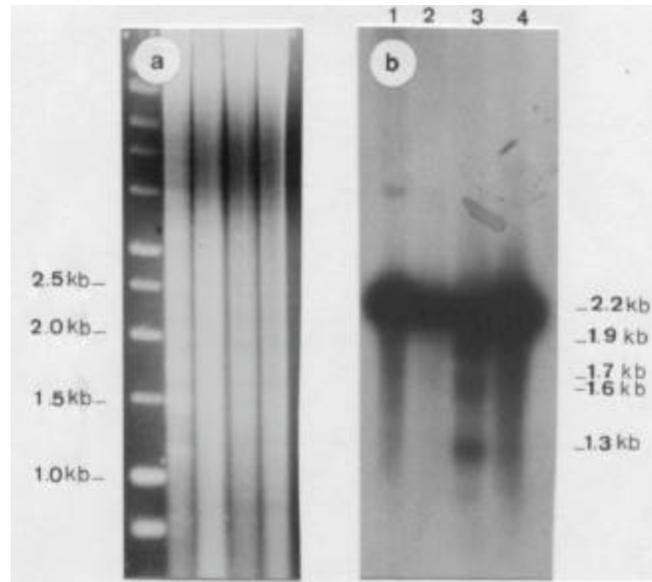
**Figure 3.**

- a. Amplification of the nuclear marker *RG64* from Taipei 309 seedlings using different concentrations of  $MgCl_2$ . Lane 1, DNA marker, lane 2, 0.5 mM  $MgCl_2$ , lane 3, 1 mM  $MgCl_2$ , lane 4, 1.5 mM  $MgCl_2$  and lane 5, 2 mM  $MgCl_2$ .
- b. The annealing temperature was raised to 50°C to remove the extra bands.
- c. Amplification of *RG64* from Taipei 309 seedlings and cells. Lane 1, DNA sample from Taipei 309 seedlings, lanes 2-4 DNA samples from 1-, 2- and 3-week-old cells.



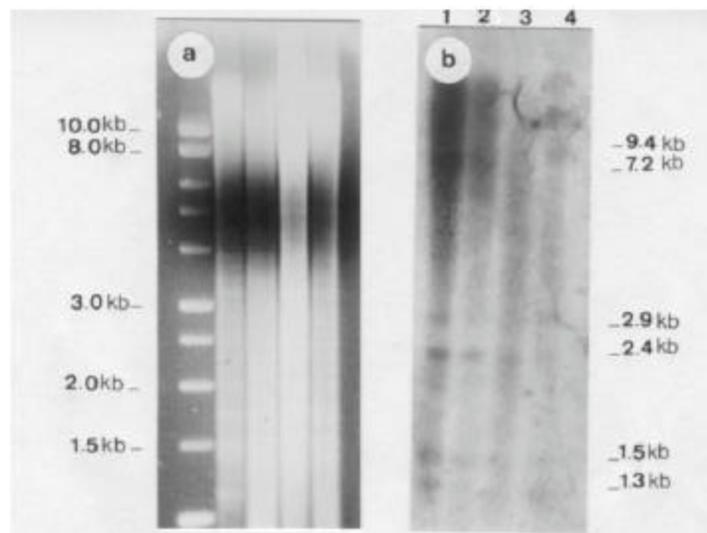
**Figure 4. Changes in mtDNA during cell senescence.**

- a. Digestion of DNA samples 5  $\mu$ g total DNA in each lane from Taipei 309 seedlings and cells with *Bam*HI. From left to right: DNA marker and DNA from seedlings, 1-, 2- and 3-week-old cells.
- b. The hybridisation of 5' end of *coxII* (the PCR product) to the *Bam*HI digested DNA samples from Taipei 309 seedlings and cells. Lane 1, DNA sample from Taipei 309 seedlings, lanes 2-4 DNA samples from 1-, 2- and 3-week-old cells.



**Figure 5. Changes in plastid DNA during cell senescence.**

- a. Digestion of DNA samples from Taipei 309 seedlings and cells with *Bam*HI. From left to right: DNA marker and DNA from seedlings, 1-, 2- and 3-week-old cells.
- b. The hybridisation of a fragment of *psaA* to *Bam*HI digested DNA samples from Taipei 309 seedlings and cells. Five  $\mu$ g of DNA was used for all samples. Lane 1, DNA sample from Taipei 309 seedlings, lanes 2-4 DNA samples from 1-, 2- and 3-week-old cells.



**Figure 6. Changes in nuclear DNA during cell senescence.**

- a. Digestion of DNA samples from Taipei 309 seedlings and cells with *Bam*HI. From left to right: DNA from seedlings, 1-, 2- and 3-week-old cells.
- b. The hybridisation of a fragment of *RG64* to *Bam*HI digested DNA samples from Taipei 309 seedlings and cells. Five  $\mu$ g of DNA was used for all samples. Lane 1, DNA sample from Taipei 309 seedlings, lanes 2-4 DNA samples from 1-, 2- and 3-week-old cells.